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1 Letter to the Editor: The *Ralstonia* research community rejects the proposal to classify
2 phylotype I *Ralstonia* into the new species *Ralstonia nicotianae*.

3
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43 The *Ralstonia solanacearum* species complex (RSSC) is a group of globally important plant pathogens.
44 Bacteria in this very large and genetically diverse group all colonize the xylem elements of angiosperm
45 plants and cause high-impact wilting diseases of many crops. Because they threaten economic and food
46 security, several RSSC subgroups are strictly regulated as quarantine pests (See “Regulation” section of
47 References). Biologically meaningful and consistent nomenclature is essential for organisms that have
48 major economic and regulatory importance like plant pathogenic *Ralstonia*. There are currently three
49 species of *Ralstonia* wilt pathogens: *R. pseudosolanacearum* (corresponding to two phylogenetic groups
50 that are described in the literature as phylotypes I and III), *R. solanacearum* (phylotypes IIA, IIB, and IIC),
51 and *R. syzygii* (phylotype IV, containing three subspecies: subsp. *syzygii*, subsp. *celebensis*, and subsp.
52 *indonesiensis*). A recent paper proposed re-classifying phylotype I as a new species named “*Ralstonia*
53 *nicotianae*” (Liu et al. 2023). The purpose of this commentary is to register our objection to the taxon
54 “*Ralstonia nicotianae*”.

55

56 **Although changing bacterial taxonomy is sometimes necessary, the *Ralstonia nicotianae***
57 **proposal is not justified.** Changing the taxonomy of any groups of organisms can be disruptive to both
58 scientists and regulators, so it should not be proposed for trivial reasons, as explained by the code of the
59 International Committee on Systematics of Prokaryotes (ICSP) (Oren et al. 2023). There are two main
60 reasons to propose new bacterial species. First, the isolation and discovery of novel bacteria that do not
61 belong to any named species justifies the naming of a new species. Second, better data from
62 technological and analytical advances can change our understanding of the diversity and evolution of
63 bacterial lineages. With sufficient evidence, these advances can justify taxonomic revisions so that the
64 newly named species better reflect evolutionary relationships. However, the *R. nicotianae* proposal is not
65 based on discovery of a new lineage, nor does it reflect novel insight into the evolutionary relationships
66 within the RSSC. As demonstrated below, the *R. nicotianae* proposal ignores natural phylogenetic gaps
67 among the existing three species. Moreover, it is based on inappropriately selective use of molecular
68 analyses.

69

70 During their decades-long careers, Drs. Philippe Prior and Mark Fegan collected and studied the diversity
71 of RSSC plant pathogens from around the world. Both research group leaders concurred that the RSSC
72 is properly divided into three species (Fegan and Prior 2005; Remenant et al. 2010, 2011; Prior et al.
73 2016; Safni et al. 2014). Specifically, extensive genomic and biological analyses of phylotype I and III
74 strains led these and other experts to conclude that phylotype I and III should not be divided into distinct

75 species (Prior et al. 2016; Truchon et al. 2023; Sharma et al. 2022). As a result, three RSSC species
76 were validly published in 2014 as *R. solanacearum*, *R. pseudosolanacearum*, and *R. syzygii* (Safni et al.
77 2014). These names were subsequently validated by the list editors of the *International Journal of*
78 *Systematic and Evolutionary Microbiology* (IJSEM). Figure 1 shows the overall relationships among
79 subgroups of the RSSC in a phylogenetic tree constructed using the core genome of the species
80 complex. *R. pseudosolanacearum* is composed of two major subgroups (phylotype I and III). *R.*
81 *solanacearum* is composed of three major subgroups (phylotype IIA, IIB, and IIC).

82

83 **Natural gaps in genetic diversity separate the three RSSC species.** Average nucleotide identity (ANI)
84 is now an accepted way to use whole genome sequences to measure relationships between strains and
85 propose species delineations (Oren et al. 2023). The ANI between any pair of genomes can be calculated
86 based on different algorithms, such as BLAST comparisons (“ANIb”) or the MUMMER index (“ANIm”). We
87 used pyani (Pritchard et al. 2016), a Python-based ANIb software, to calculate pairwise ANIb values for
88 300 RSSC genomes, including genomes of 11 phylotype III strains and 148 phylotype I strains. When the
89 resulting 90,000 ANIb values are hierarchically clustered and visualized as a heatmap, three obvious
90 clusters correspond to the three accepted RSSC species (Fig. 2A).

91

92 **An ANI threshold of 96% is not the appropriate cut-off for delineating species in the RSSC.**

93 Depending on the taxon, bacterial species borders can be drawn using ANI threshold values of 95-96%
94 (Oren et al. 2023; Chun et al. 2018). However, ANI <95% is the most widely used cut-off for dividing
95 species. This threshold has been applied across the bacterial domain by the Genome Taxonomy
96 Database (Parks et al. 2020). We investigated the distributions of 90,000 ANIb comparisons among 300
97 RSSC genomes to determine if there is a biologically relevant cut-off that separates RSSC species.

98

99 The *R. nicotianae* proposal applied an ANI = 96% species threshold value. However, our analysis of 300
100 RSSC genomes suggests that 95% is the appropriate threshold for delineating species within the RSSC
101 (Fig. 2). Visualizing the distribution of ANI values reveals an obvious natural gap in ANIb values: no
102 pairwise comparison yields an ANI value between 92.57% and 95.06% (Fig. 2B). Applying an ANI cut-off
103 of 96% (indicated by the red lines in Figure 2 graphs) would interrupt a continuous distribution of genetic
104 distances within the RSSC as a whole (Fig. 2B), within *R. solanacearum* (Fig. 2C) and within *R.*
105 *pseudosolanacearum* (Fig. 2D). In contrast, an ANI = 95% cut-off (indicated by the blue lines) separates
106 the RSSC into three species with clear gaps that suggest that these groups have distinct evolutionary

107 histories (Fig. 2E) and the existing 3-species nomenclature may thus represent their natural phylogenetic
108 order.

109

110 **The data presented in the *R. nicotianae* proposal do not support a division of phylotype I into a**
111 **new species.** This section provides a detailed dissection of ANI data to highlight the methodological
112 problems in the *R. nicotianae* proposal.

113

114 The *R. nicotianae* proposal was based on limited analyses that compared genomes of a single phylotype
115 I and a single phylotype III genome against other *R. pseudosolanacearum* genomes. This approach
116 significantly biased the statistics and phylogenetic analyses, as it does not reflect the diversity of a
117 representative population of isolates. The focal strains were the established type strain of *R.*
118 *pseudosolanacearum* (phylotype III strain LMG9673^T) and a phylotype I strain (RS) that was proposed as
119 a type strain for the novel species. Hereafter, we refer to this strain as RS^{proposed_T}.

120

121 The *R. nicotianae* proposal calculated ANI with three methods: FastANI using the Genome Taxonomy
122 Database website interface, ANIb using the JSpeciesWS website interface, and MUMMER-based ANI
123 (ANIm) using the JSpeciesWS website interface. The authors then carried out 434 FastANI comparisons
124 (LMG9673^T and RS^{proposed_T} vs. 204 phylotype I and 11 phylotype III strains), 24 ANIb comparisons
125 (LMG9673^T and RS^{proposed_T} vs. 1 phylotype I and 11 phylotype III strains), and 24 ANIm comparisons
126 (LMG9673^T and RS^{proposed_T} vs. 1 phylotype I and 11 phylotype III strains).

127

128 Comparing RS^{proposed_T} to the 11 phylotype III genomes yielded FastANI values from 95.85 to 96.06%,
129 ANIm values from 96.12 to 96.26%, and ANIb values from 94.95 to 95.33% as described in the *R.*
130 *nicotianae* proposal. We also computed ANIb values, but we used the Python-based pyani tool over a
131 larger sample size of phylotype I and III genomes (Fig. 2). In the subset of comparisons that overlap
132 between our analysis and that of the *R. nicotianae* proposal, pyani yielded ANIb values from 95.77-
133 96.02%. An overview of these data are presented in Fig. 3A, which compares the ANI values obtained for
134 each of the comparisons and methods.

135

136 For taxonomic classification, the most important ANI comparisons are between type strains. In the *R.*
137 *nicotianae* proposal, comparisons between RS^{proposed_T} and the *R. pseudosolanacearum* type strain

138 LMG9673^T yielded values of 95.97 to 96.02% (FastANI), 96.14 to 96.15% (ANIm), and 95.23 to 95.30%
139 (ANlb). Our pyani calculation of ANlb yielded a narrow range of values from 95.81 to 95.82%.

140
141 Before genome sequences were readily available, the gold standard for classifying bacterial strains into
142 species was a wet-lab technique called DNA-DNA Hybridization (DDH). A 70% DDH threshold was used
143 to delineate bacterial species. The *R. nicotianae* proposal used three digital DDH calculations (dDDH) to
144 estimate DDH between RS^{proposed_T} and LMG9673^T. Two dDDH calculations yielded values above the
145 standard 70% threshold (74.9% and 75.8%) while a third dDDH calculation yielded a value of 66.2%. If
146 averaged, the three calculations yield 72.3%, above the 70% species cut-off. Fig. 3B shows the full
147 distribution of dDDH scores from the *R. nicotianae* proposal. However, the text of the *R. nicotianae*
148 proposal emphasized only the lowest of these three DDH values.

149
150 The careful assessment above reveals that the conclusions in the *R. nicotianae* proposal were based on
151 the sole DDH analysis and the sole ANI analysis where comparisons of type strains yielded a value less
152 than the 70% DDH threshold and an ANI value in the gray zone of 95-96% ANI. This ignored the
153 molecular phylogenomic analysis results that suggested that phylotype I should remain within the *R.*
154 *pseudosolanacearum* species. Selecting among obtained results to present only the subset of results that
155 support a preferred narrative is not consistent with good scientific practice (Casadevall and Fang 2016).

156
157 **Even if there was genomic and biological justification for the separation of phylotype I into a novel**
158 **species, *nicotianae* would be a misleading species epithet for phylotype I.** The epithet *nicotianae*
159 was suggested because pathogenic bacteria are sometimes named for their host of isolation, usually the
160 primary host, and the proposed Type strain RS^{proposed_T} was isolated from an experimental tobacco plot.
161 However, this name would be misleading because infecting tobacco is not a distinguishing trait of
162 phylotype I. RSSC strains from each of the four phylotypes have been isolated from tobacco (Lowe-
163 Power et al. 2020). Furthermore, phylotype I strains have the broadest host range within the RSSC;
164 phylotype I strains have been isolated from 95 plant species in 79 genera in 46 families (Lowe-Power et
165 al. 2020). In comparison, the other three phylotypes combined have been isolated from only 69 plant
166 species in 40 genera in 28 families (Lowe-Power et al. 2020).

167
168 Proposing new names without careful consideration can create confusion in the research community and
169 potentially in the published literature. For example, the widely-used NCBI genome database transiently

170 adopted the *R. nicotianae* proposal. Within two weeks of the publishing of the *R. nicotianae* proposal in
171 *Frontiers in Microbiology*, we noticed that NCBI had renamed the genome of the much studied model *R.*
172 *pseudosolanacearum* strain GMI1000 to “*Ralstonia nicotianae*”. This occurred before the IJSEM list
173 editors had the opportunity to consider this proposal and issue a decision about publishing the new name.
174 Although GMI1000 is a phylotype I *R. pseudosolanacearum* strain, the GMI1000 genome was still labeled
175 in NCBI as “*Ralstonia solanacearum*” for historical reasons: the genome was sequenced and deposited
176 14 years before the RSSC was formally divided into three species (Salanoubat et al. 2002). Importantly,
177 this error was promptly corrected when it was brought to the attention of NCBI.

178
179 **Summary.** Adopting “*R. nicotianae*” as a newly named species corresponding to phylotype I and reducing
180 the validly published species *R. pseudosolanacearum* to include only phylotype III is not justified based
181 on either genomic similarity or evolutionary relationships. On the contrary, the comparative genomics
182 analyses presented in the *R. nicotianae* proposal are consistent with the conclusion that phylotype I and
183 phylotype III are two subgroups of the same species, *R. pseudosolanacearum*. Furthermore, accepting a
184 division of phylotype I and III into separate species would complicate and disrupt scientific and regulatory
185 communication about strains and genomes of plant pathogenic *Ralstonia*. Changing the name of a taxon
186 that has been established and validated through multiple rigorous studies would create unnecessary
187 confusion. This proposal violates three of the four essential elements of Principle 1 of the International
188 Code of Nomenclature of Prokaryotes, which states that nomenclature should: “1) Aim at stability of
189 names; 2) Avoid or reject names that create error or confusions; and 3) Avoid the useless creation of
190 names” (Oren et al. 2023). Finally, the chosen species name would be misleading regarding the host
191 range of the strains that belong to it and to the related strains in other species within the RSSC, and is
192 thus in conflict with International Code of Nomenclature of Prokaryotes Recommendation 12(c) 2: “Avoid
193 [epithets] that express a character common to all, or nearly all, the species of a genus” (Oren et al.
194 2023). These reasons, together with the analyses presented in this letter, establish that “*Ralstonia*
195 *nicotianae*” Liu et al. 2023 is at most a junior heterotypic synonym of *Ralstonia pseudosolanacearum*
196 Safni et al. 2014.

197
198 Therefore, we strongly encourage our fellow scientists in the RSSC community not to adopt *R. nicotianae*
199 in publications and scientific communication in general. We further respectfully request that the IJSEM list
200 editors review the evidence presented here when considering whether *R. nicotianae* should be validly
201 published.

202

203 **References**

- 204 Bayliss, S. C., Thorpe, H. A., Coyle, N. M., Sheppard, S. K., and Feil, E. J. 2019. PIRATE: A fast and
205 scalable pangenomics toolbox for clustering diverged orthologues in bacteria. *Gigascience*. 8(10):
206 giz119.
- 207 Casadevall, A., and Fang, F. C. 2016. *Rigorous Science: a How-To Guide*. MBio. 7 Available at:
208 <http://dx.doi.org/10.1128/mBio.01902-16>.
- 209 Chun, J., Oren, A., Ventosa, A., Christensen, H., Arahal, D. R., da Costa, M. S., et al. 2018. Proposed
210 minimal standards for the use of genome data for the taxonomy of prokaryotes. *Int. J. Syst. Evol.*
211 *Microbiol.* 68:461–466.
- 212 Fegan, M., and Prior, P. 2005. How complex is the *Ralstonia solanacearum* species complex? In
213 *Bacterial wilt disease and the Ralstonia solanacearum species complex*, APS press St. Paul, p.
214 449–461.
- 215 Liu, J. Y., Zhang, J. F., Wu, H. L., Chen, Z., Li, S. Y., Li, H. M., et al. 2023. Proposal to classify *Ralstonia*
216 *solanacearum* phylotype I strains as *Ralstonia nicotianae* sp. nov., and a genomic comparison
217 between members of the genus *Ralstonia*. *Front. Microbiol.* 14:1135872.
- 218 Lowe-Power, T., Avalos, J., Bai, Y., Charco Munoz, M., Chipman, K., Tom, C., and Williams, D. 2022. A
219 meta-analysis of the known global distribution and host range of the *Ralstonia* species complex.
220 bioRxiv. doi.org/10.1101/2020.07.13.189936
- 221 Minh, B. Q., Schmidt, H. A., Chernomor, O., Schrempf, D., Woodhams, M. D., von Haeseler, A., et al.
222 2020. IQ-TREE 2: New models and efficient methods for phylogenetic inference in the genomic
223 era. *Mol. Biol. Evol.* 37:1530–1534.
- 224 Oren A, Arahal DA, Göker M, Moore ERB, Rossello-Mora R, Sutcliffe IC (eds). 2023. International code of
225 nomenclature of prokaryotes. Prokaryotic Code (2022 Revision). *Int J Syst Evol Microbiol*, in
226 press.
- 227 Parks, D. H., Chuvochina, M., Chaumeil, P.-A., Rinke, C., Mussig, A. J., and Hugenholtz, P. 2020. A
228 complete domain-to-species taxonomy for Bacteria and Archaea. *Nat. Biotechnol.* 38:1079–1086.
- 229 Prior, P., Ailloud, F., Dalsing, B. L., Remenant, B., Sanchez, B., and Allen, C. 2016. Genomic and
230 proteomic evidence supporting the division of the plant pathogen *Ralstonia solanacearum* into
231 three species. *BMC Genomics.* 17:90.

- 232 Pritchard, L., Glover, R. H., Humphris, S., Elphinstone, J. G., and Toth, I. K. 2016. Genomics and
 233 taxonomy in diagnostics for food security: soft-rotting enterobacterial plant pathogens. *Anal.*
 234 *Methods.* 8:12–24.
- 235 Remenant, B., de Cambiaire, J.-C., Cellier, G., Jacobs, J. M., Mangenot, S., Barbe, V., et al. 2011.
 236 *Ralstonia syzygii*, the Blood Disease Bacterium and some Asian *R. solanacearum* strains form a
 237 single genomic species despite divergent lifestyles. *PLoS One.* 6:e24356.
- 238 Remenant, B., Coupat-Goutaland, B., Guidot, A., Cellier, G., Wicker, E., Allen, C., et al. 2010. Genomes
 239 of three tomato pathogens within the *Ralstonia solanacearum* species complex reveal significant
 240 evolutionary divergence. *BMC Genomics.* 11:379.
- 241 Safni, I., Cleenwerck, I., De Vos, P., Fegan, M., Sly, L., and Kappler, U. 2014. Polyphasic taxonomic
 242 revision of the *Ralstonia solanacearum* species complex: proposal to emend the descriptions of
 243 *Ralstonia solanacearum* and *Ralstonia syzygii* and reclassify current *R. syzygii* strains as
 244 *Ralstonia syzygii* subsp. *syzygii* subsp. nov., *R. solanacearum* phylotype IV strains as *Ralstonia*
 245 *syzygii* subsp. *indonesiensis* subsp. nov., banana blood disease bacterium strains as *Ralstonia*
 246 *syzygii* subsp. *celebesensis* subsp. nov. and *R. solanacearum* phylotype I and III strains as
 247 *Ralstonia pseudosolanacearum* sp. nov. *Int. J. Syst. Evol. Microbiol.* 64 :3087-3103.
- 248 Salanoubat, M., Genin, S., Artiguenave, F., Gouzy, J., Mangenot, S., Arlat, M., et al. 2002. Genome
 249 sequence of the plant pathogen *Ralstonia solanacearum*. *Nature.* 415:497–502.
- 250 Sharma, P., Johnson, M. A., Mazloom, R., Allen, C., Heath, L. S., Lowe-Power, T. M., et al. 2022. Meta-
 251 analysis of the *Ralstonia solanacearum* species complex (RSSC) based on comparative
 252 evolutionary genomics and reverse ecology. *Microb Genom.* 8:00079.
- 253 Truchon, A. N., Dalsing, B. L., Khokhani, D., MacIntyre, A., McDonald, B. R., Ailloud, F., et al. 2023.
 254 Plant-pathogenic *Ralstonia* phylotypes evolved divergent respiratory strategies and behaviors to
 255 thrive in xylem. *MBio.* 14:e0318822.

256

257 **Regulation:**

- 258 European Union: Regulation (EU) 2016/2031 of the European Parliament of the Council of 26 October
 259 2016 on protective measures against pests of plants, amending Regulations (EU) No 228/2013,
 260 (EU) No 652/2014 and (EU) No 1143/2014 of the European Parliament and of the Council and
 261 repealing Council Directives 69/464/EEC, 74/647/EEC, 93/85/EEC, 98/57/EC, 2000/29/EC,
 262 2006/91/EC and 2007/33/EC <https://eur-lex.europa.eu/eli/reg/2016/2031/oj>

263 United Kingdom: [https://planthealthportal.defra.gov.uk/pests-and-diseases/pest-and-disease-](https://planthealthportal.defra.gov.uk/pests-and-diseases/pest-and-disease-factsheets/notifiable-diseases/)
 264 [factsheets/notifiable-diseases/](https://planthealthportal.defra.gov.uk/pests-and-diseases/pest-and-disease-factsheets/notifiable-diseases/)
 265 Canada: [https://inspection.canada.ca/plant-health/invasive-species/regulated-](https://inspection.canada.ca/plant-health/invasive-species/regulated-pests/eng/1363317115207/1363317187811)
 266 [pests/eng/1363317115207/1363317187811](https://inspection.canada.ca/plant-health/invasive-species/regulated-pests/eng/1363317115207/1363317187811)
 267 United States: <https://www.aphis.usda.gov/aphis/ourfocus/planthealth/import-information/rppl/rppl-table>
 268
 269

270 **Fig. 1. Core genome phylogenetic tree demarcating the three RSSC species** and their major
 271 subdivisions: *R. pseudosolanacearum* (phylotype I and III subdivisions), *R. solanacearum* (phylotype IIA,
 272 IIB, and IIC subdivisions (Sharma et al. 2022)), and *R. syzygii*. Tree was built using IQtree (Minh et al.
 273 2020) using the core-genome alignments obtained with PIRATE (Bayliss et al. 2019) as input.

274
 275 **Fig. 2. The biologically relevant ANI threshold for delineating RSSC species is 95%.** (A) Robust ANI
 276 analysis of 300 RSSC genomes reveals three species clusters corresponding to *R. pseudosolanacearum*,
 277 *R. solanacearum*, and *R. syzygii*. Pairwise comparisons are shown in an ANI heatmap calculated with the
 278 BLAST-based ANIb method using pyani (Pritchard et al. 2016). (B) The distribution of pairwise ANIb
 279 values between 300 RSSC strains reveals a natural gap between pairs sharing 92.57% and 95.06%
 280 ANIb. ANIb was calculated with pyani (Pritchard et al. 2016). (C) Comparison of ANI values within the *R.*
 281 *pseudosolanacearum* species and its two major subdivisions. (D) Comparison of ANI values within the *R.*
 282 *solanacearum* species and its three major subdivisions. (E) Comparison of ANI values between the three
 283 validated RSSC species. Blue lines show the biologically relevant ANI threshold of 95% and red lines
 284 show the biologically inappropriate threshold of 96%.

285
 286 **Fig 3. The *R. nicotianae* proposal focused on outlier ANI and dDDH calculations that supported a**
 287 **new species.** (A) Comparison of ANI values from the 12 pairs of genomes that were shared between the
 288 *R. nicotianae* proposal and our larger-scale analysis (Figs 1 and 2). The *R. nicotianae* proposal analyzed
 289 ANI between six phylotype III genomes to two strains: the *R. pseudosolanacearum* type strain
 290 (LMG9673^T) and the phyl. I strain proposed as a new type strain (RS^{proposed-T}). (B) Comparison of dDDH
 291 calculations from the *R. nicotianae* proposal. Lines connect the same strain pairings that were analyzed
 292 using three different dDDH tools. ANI and DDH comparisons of *R. pseudosolanacearum* type strain
 293 LMG9673^T and RS^{proposed-T} are shown in red. Arrows indicate the outlier results favored in the *R.*
 294 *nicotianae* proposal.

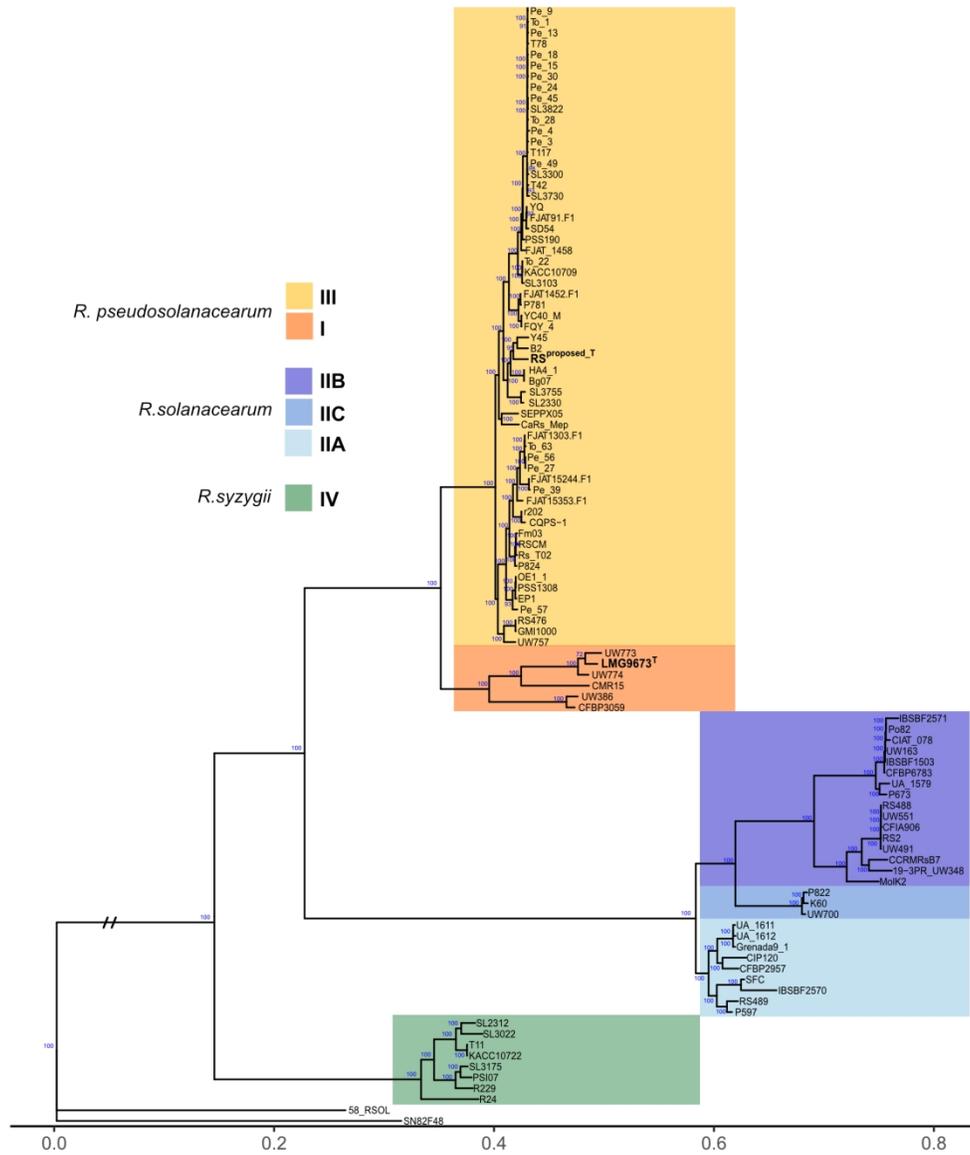


Fig. 1. Core genome phylogenetic tree demarcating the three RSSC species and their major subdivisions: *R. pseudosolanacearum* (phylogroup I and III subdivisions), *R. solanacearum* (phylogroup IIA, IIB, and IIC subdivisions (Sharma et al. 2022)), and *R. syzygii*. Tree was built using IQtree (Minh et al. 2020) using the core-genome alignments obtained with PIRATE (Bayliss et al. 2019) as input.

705x845mm (72 x 72 DPI)

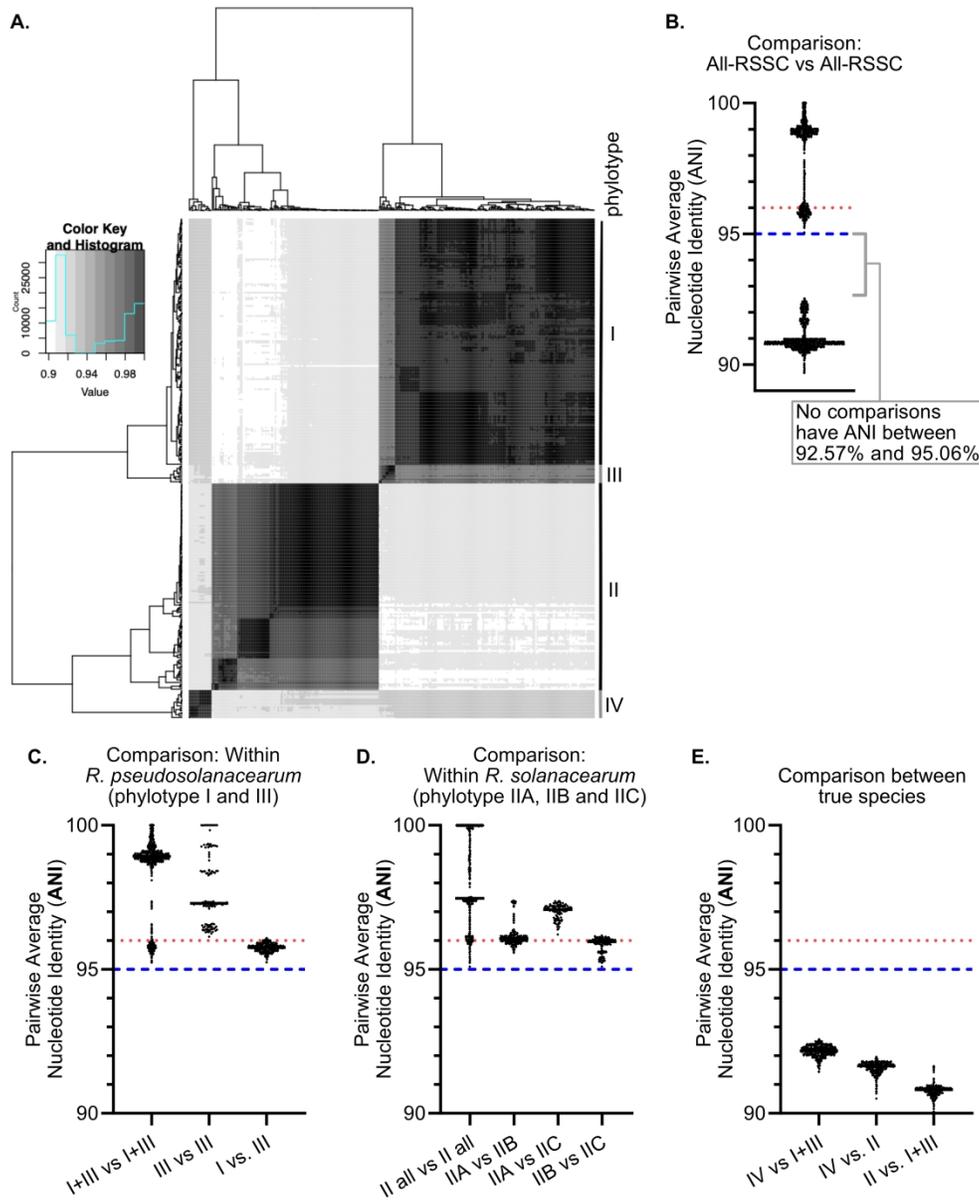


Fig. 2. The biologically relevant ANI threshold for delineating RSSC species is 95%. (A) Robust ANI analysis of 300 RSSC genomes reveals three species clusters corresponding to *R. pseudosolanacearum*, *R. solanacearum*, and *R. syzygii*. Pairwise comparisons are shown in an ANI heatmap calculated with the BLAST-based ANIb method using pyani (Pritchard et al. 2016). (B) The distribution of pairwise ANIb values between 300 RSSC strains reveals a natural gap between pairs sharing 92.57% and 95.06% ANIb. ANIb was calculated with pyani (Pritchard et al. 2016). (C) Comparison of ANI values within the *R. pseudosolanacearum* species and its two major subdivisions. (D) Comparison of ANI values within the *R. solanacearum* species and its three major subdivisions. (E) Comparison of ANI values between the three validated RSSC species. Blue lines show the biologically relevant ANI threshold of 95% and red lines show the biologically inappropriate threshold of 96%.

89x108mm (1200 x 1200 DPI)

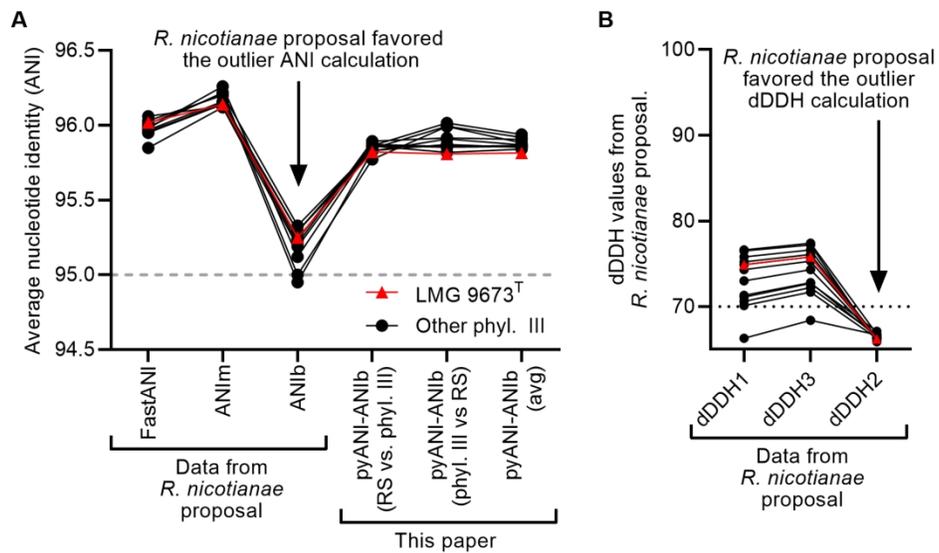


Fig 3. The *R. nicotianae* proposal focused on outlier ANI and dDDH calculations that supported a new species. (A) Comparison of ANI values from the 12 pairs of genomes that were shared between the *R. nicotianae* proposal and our larger-scale analysis (Figs 1 and 2). The *R. nicotianae* proposal analyzed ANI between six phylotype III genomes to two strains: the *R. pseudosolanacearum* type strain (LMG9673T) and the phyl. I strain proposed as a new type strain (RSproposed_T). (B) Comparison of dDDH calculations from the *R. nicotianae* proposal. Lines connect the same strain pairings that were analyzed using three different dDDH tools. ANI and DDH comparisons of *R. pseudosolanacearum* type strain LMG9673T and RSproposed_T are shown in red. Arrows indicate the outlier results favored in the *R. nicotianae* proposal.

168x103mm (300 x 300 DPI)